



Systematic study on some Urticaceae Juss. species from Egypt

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Abstract

Five out of six species belonging to the three genera of Urticaceae were collected from different location in Egypt and subjected in this study. Pollen and seed external morphological characters beside stem and lamina internal characters were examined and photographed using both Light (LM) and Scanning Electron (SEM) Microscopes to assess the use of the studied characters in the separation and identification of species. The most diagnostic characters of both stem and lamina were; dissected or continuous siphonostelic structure of the vascular tissue, dorsiventral lamina, cystolith, stinging hairs, ruminant, rugose or colliculate epidermal cell surface. As well as the shape, size, aperture type and the exine ornamentation of the pollen grains, in addition to seed surface ornamentations. These characters can be of help in the discrimination of the studied species. The most obvious variable characters obtained in constructing an artificial taxonomic key for the studied species. All the 37 characters were subjected to the classical cluster analysis (UPGMA) and the principal component analysis (PCA) using PAST version 3.16 software and clearly revealed the splitting of the studied taxa into two clusters and three groups.

Key words: Anatomy, cluster analysis, pollen, seed, Urticaceae

Introduction

Family Urticaceae Juss. (Nettle family) comprises 53 genera and 2625 species worldwide (Christenhusz & Byng 2016). Members of the family are herbs, shrubs, lianas or small trees and have a subcosmopolitan distribution with most genera and species found in the moist tropics and temperate regions. The taxonomic position of the family has been faced with many opinions and the history of the taxonomic opinions of the family is summarized in table 1. The family was described by Antoine-Laurent de Jussieu in 1789 under the name “Urticae” and divided its members under three groups based on inflorescence morphology. Gaudichaud (1830) revised the family as “La famille des urticées” and classified the genera into five tribes. Subsequent studies on the family

accepted its division into three subfamilies with five tribes (Berg 1977, 1989, Friis 1989b, 1993). Takhtajan (2009) considered it as one of the Urticales along with Ulmaceae, Moraceae, Cannabaceae, Cecropiaceae, while Stevens (2001 onwards) and the recent taxonomic works beside the APG III system (2009) placed it in order Rosales closer to Rosaceae.

The research studies on the morphology of Urticaceae are few, in spite of that we can refer to Gangadhara & Inamdar (1977), Lersten & Curtis (1991), Abid et al. (2015) and Hamdy et al. (2016). Members within the Urticaceae show remarkable morphological diversity; the plants are monoecious, dioecious rarely polygamous. The leaves simple, sessile or petiolate with alternate or opposite arrangement.

Table 1. Taxonomic opinions of the family Urticaceae.

Author	Subfamily	Tribes	Studied genera
Gaudichaud (1830)		Urereae	<i>Urtica</i>
		Elatostemeae	
		Boehmerieae	
		Parietarieae	<i>Parietaria</i>
Weddell (1869)		Forskalieae	<i>Forsskaolea</i>
		Urereae	<i>Urtica</i>
		Procridaeae	
		Boehmerieae	
Friis (1993)		Parietarieae	<i>Parietaria</i>
		Forskalieae	<i>Forsskaolea</i>
		Urticeae	<i>Urtica</i>
		Lecantheae	
Kravtsova (2009)		Boehmerieae	
		Parietarieae	<i>Parietaria</i>
		Forskalieae	<i>Forsskaolea</i>
		Urticeae	<i>Urtica</i>
APG III (2009)	Urticoideae		
	Lecantheoideae	Lecantheae	
	Boehmerioideae	Boehmerieae	
		Parietarieae	<i>Parietaria</i>
		Forskalieae	<i>Forsskaolea</i>
		Boehmerieae	
APG III (2009)		Cecropieae	
		Elaostemateae	
		Forsskaoleae	<i>Forsskaolea</i>
		Parietarieae	<i>Parietaria</i>
		Urticeae	<i>Urtica</i>

Ducts and internal characters are more interesting within the family, Guerin (1923) studied the mucilaginous cells and lactiferous ducts in the vegetative parts of some taxa and reported the mucilaginous cells in *Urtica*, *Nanocnide*, *Laportea*, *Girardinia*, *Urera* and *Gyrotaenia* and the lactiferous ducts were observed in *Urera*. Friis & Wilmot- Dear (1988) investigated and revised tribe Forsskaoleae in which they maintained the two tribes within the tribe based on the inflorescence and floral structure. Metcalfe & Chalk (1950) reported that the cortex is well developed in stem section with stone cells in *Urera* but not in other genera, the pericyclic fibers present in the form of

isolated strands and the xylem in the form of continuous cylinder traversed by narrow rays in some species. They also reported dorsiventral lamina; glandular, e-glandular, stinging trichomes and anomocytic stomata. Jafari & Dehghan (2012) studied the anatomical structure of aerial organs in four populations of *Urtica dioica*. Hamdy *et al.* (2016) studied the anatomical features of stem and leaf of *Forsskaolea tenacissima* growing in Egypt. Pollen grain morphology done by Sorsa & Huttunen (1975), they found that the structure, size of the pores and exine sculpture were considered the most diagnostic pollen characteristics. Friis (1989

Systematic study on some Urticaceae Juss. species from Egypt

b) reported that the pollen grains of the family are small sized, oblate to suboblate-spheroidal, porate (two to six pores) or foraminate (up to 15), with more or less obscure exine sculpture. He observed that in tribe Urticeae the pollen triporate, sometimes polyporate (four to seven pore) with small sized grains varies from 10- 29 μm while in *Nanocnide*, *Urera* and *Hesperocnide* the pores are wider, more regularly and densely placed spinules exine sculpture.

The seed morphological characters were significant and useful in revealing the taxonomic relationship between the species of flowering plants (Barthlott 1981; Ather *et al.* 2010; Kanwal *et al.* 2012; El-Ghamery *et al.* 2018). The seed morphology of Urticaceae has been studied by some researchers (Chen *et al.* 2003; Abid *et al.* 2015).

In Egypt, this family is represented by seven species belonging to three genera *viz.* *Forsskaolea* L., *Parietaria* L. and *Urtica* L. (Täckholm 1974), while Boulos (1999) enumerated six species. The specific objective of the present study is the investigation of the morphological characters of stems, lamina, pollen grains and seeds

(macro- and micro-characters) to evaluate the weight of these characters in the delimitation and identification between the studied species of Urticaceae growing in Egypt. As well as to realize the nearest similar taxa and their differentiations and recognition by constructing artificial key for the studied species.

Materials:

In the present study, field trips have been done to collect five out of six species from their natural habitats in Egypt (Table 2, map 1). Ten individuals from each location have subjected for this investigation. Pieces from the fourth stem internodes and leaf lamina were taken and mounted in F.A.A. solution for sectioning. Samples of flowers and seeds have been collected for laboratory investigations. The identification and synonyms of the studied species followed Täckholm (1974), Boulos (1999) and International Plant Names Index (IPNI). Sample individuals were prepared for herbarium preservation in Botany Department, Faculty of Science, Ain Shams University herbarium (CAIA).

Table 2. Data collection

No.	Species	Locality	Life span / date of collection
1	<i>Forsskaolea tenacissima</i> L. = <i>F. cossoniana</i> Webb = <i>Caidbeja adhaerens</i> Forssk.	South Sinai Governorate (Wadi Feiran)	Perennial herb 5 / 2019
2	<i>Parietaria alsinifolia</i> Delile = <i>Freirea alsinaefolia</i> (Delile) Gaudich.	South Sinai Governorate, Saint Katherine protectorate (Wadi El-Arbaeen)	Perennial herb 5 / 2019
3	<i>P. judaica</i> L. = <i>P. diffusa</i> Mert. & W. D. J. Koch = <i>P. punctata</i> Willd.	Botanical garden, Faculty of Science, Ain Shams University	Perennial herb 6 / 2019
4	<i>Urtica pilulifera</i> L.	Kafr El- Sheikh Governorate, around cultivated lands	Annual herb 3/ 2019
5	<i>U. urens</i> L.	Mediterranean Coastal Region, Burg El-Arab (Brimley cave)	Annual herb 3/ 2019



Map 1. Different locations of the studied species

Methods:

A- Stem and lamina anatomy

Transverse sections in the fourth internodes of stem and vertical sections in the fourth laminae were done for LM investigation and photography. Sectioning in the studied specimens done by hand microtome at 10-15 μm thick. The sections were double stained using safranin (2%) and light green (1%), and finally mounted in Canada Balsam according to the conventional method (Johansen 1940). Sections were examined and photographed using Olympus C.35AD-2. The terminology of the internal structures followed Metcalfe & Chalk (1950).

B- Lamina epidermal investigation

Lamina epidermal peels were taken from abaxial surface using forceps and mounted on glass slide directly for light microscope. Examination and photographs were taken using Canon power-shot A470, 7.1 mega pixels. For SEM investigation, small pieces of the cleaned and dried leaves were fixed directly onto EM stubs with double-sided adhesive tape at abaxial surface and coated

with 30 nm gold in a DII-29030SCTR Smart Coater, then scanned and photographed using JEOL JSMIT100 SEM at SEM lab, the institute of Nanoscience and Nanotechnology, Kafr El- Sheikh University. Terminology of epidermal cell features followed Stace (1984) and Prabhakar (2004).

C- Pollen grain morphology

Mature anthers from the collected flowers were carefully opened using sharp needles and sputtered onto glass slides. The pollen grains were examined, measured and photographed using Canon power-shot A470, 7.1 mega pixels. At least ten pollen grains / taxa were measured by LM. Polar axis (P), equatorial diameter (E) and exine thickness were measured. Other pollen characters *viz.*, size, shape and aperture type were recorded. The arithmetic mean value was calculated and the terminology used for describing pollen grains morphology followed Erdtman (1952), Punt et al. (2007) and Hesse et al. (2009). For SEM investigation, non-acetolyzed pollen grains sputtered onto EM

Systematic study on some Urticaceae Juss. species from Egypt

stubs using double-sided cello tape, coated with 30 nm gold then scanned and photographed using JEOL JSM-IT100 SEM for exine and aperture ornamentations. The terminology used here followed Erdtman (1952).

D- Seed morphology

Mature seeds (10 - 15 seed from each taxa) were examined using stereo-microscope and photographed by inserting Canon power-shot A470, 7.1 mega pixels camera on the stereomicroscope. Stage micrometer in addition to image J software was used for seed measurements and calibration. Seed colour, length and width, as well as hilum position were studied by stereomicroscope. For SEM investigation, the mature seeds were mounted onto SM stubs, coated with 30 nm gold and examined and photographed using JEOL JSM-IT100 SEM. The terminology used followed Barthlott (1981) and Stearn (1992).

E- Numerical analysis

The studied characters were scored, coded and numerically analyzed using PAST program version 3.16. The principal component analysis (PCA) was performed to assess the degree of similarity inside data matrix by un-weighted pair-group method (UPGMA) generating a dendrogram to detect the relationship between the studied species.

Results

The morphological characters of the studied species including stem and lamina anatomical characters, lamina epidermal characters, pollen characters as well as seed characters (LM & SEM) were summarized in tables 3 - 6. Some of the specific structures (micro-photographs) were arranged and illustrated in plates I - IV.

A- Stem and lamina anatomy (Table 3, Plate I)

Stem ridged and furrowed in *Urtica pilulifera* and *U. urens* or terete in *Forsskaolea tenacissima*, *Parietaria alsinifolia* and *P. judaica*. Epidermis radially elongated in *Forsskaolea tenacissima* and *Urtica pilulifera* or tangentially elongated *Parietaria alsinifolia*, *P. judaica* and *U. urens*. Cuticle thin in *Parietaria alsinifolia* and *P. judaica* or thick in *Forsskaolea tenacissima*, *Urtica pilulifera* and *U. urens*. In all the studied species, trichomes eglandular unicellular. The cortex of polyhedral parenchyma in *Parietaria alsinifolia*, polyhedral parenchyma and angular collenchyma in *Urtica pilulifera*, polyhedral parenchyma, angular collenchyma and chlorenchyma in *U. urens* or polyhedral parenchyma, angular collenchyma and extraxylary fibers in *Forsskaolea tenacissima* and *P. judaica*. The vascular supply is in the form of dissected siphonostelic structure in *Urtica pilulifera* and *U. urens* or continuous siphonostelic structure *Forsskaolea tenacissima*, *Parietaria alsinifolia* and *P. judaica*. The xylem at the interfascicular regions is of parenchyma in *Urtica pilulifera* and *U. urens* or fibers in *Forsskaolea tenacissima*, *Parietaria alsinifolia* and *P. judaica*. Stem hollow in *Parietaria alsinifolia* and *Urtica pilulifera* or solid *Forsskaolea tenacissima*, *P. judaica* and *U. urens*.

The outline of the lamina is flattened, dorsiventral and differentiated into midrib and two wings in all studied species. The epidermis is tangentially elongated in *Parietaria alsinifolia* and *Urtica urens* or radially elongated in *Forsskaolea tenacissima*, *P. judaica* and *U. pilulifera*. The mesophyll is differentiated into compact palisade parenchyma beneath upper epidermis and spongy parenchymatous tissue. Cystolith detected in *Forsskaolea tenacissima* and *Urtica urens* or wanting in *Parietaria alsinifolia*, *P. judaica* and *U. pilulifera*. In midrib region, the mechanical

tissue (angular collenchyma) detected adaxially in *Parietaria judaica* and *Urtica pilulifera* or wanting in *Forsskaolea tenacissima*, *P. alsinifolia* and *U. urens*. The vascular bundles three in *Urtica pilulifera* or represented by one mass in *Forsskaolea tenacissima*, *Parietaria alsinifolia*, *P. judaica* and *U. urens*.

B- Lamina epidermal investigation (abaxial surface; Table 4, Plate II)

In all the studied species the shapes of the epidermal cell are irregular with anomocytic stomata and covered by e-glandular unicellular hairs, except in *Urtica pilulifera* and *U. urens* where the glandular stinging hairs recorded in addition to e-glandular ones. The anticlinal walls sinuate in *Parietaria alsinifolia* and *P. judaica* or undulate in *Forsskaolea tenacissima*, *Urtica pilulifera* and *U. urens*. Sand crystals absent in *Forsskaolea tenacissima* and *Parietaria judaica* or present in *P. alsinifolia*, *Urtica pilulifera* and *U. urens*. The lamina abaxial surface sculpture rugose in *Parietaria alsinifolia*, ruminant in *Forsskaolea tenacissima* and *P. judaica* or colliculate in *Urtica pilulifera* and *U. urens*. The anticlinal wall smooth and narrow in all studied species; raised in *Forsskaolea tenacissima* and *Parietaria judaica* or sunken in *P. alsinifolia*, *Urtica pilulifera* and *U. urens*. The periclinal wall sunken in *Forsskaolea tenacissima* and *Parietaria judaica* or raised in *P. alsinifolia*, *Urtica pilulifera* and *U. urens*. The periclinal wall surface granulate in *Forsskaolea tenacissima*, striate in *Parietaria alsinifolia* or smooth in *P. judaica*, *Urtica pilulifera* and *U. urens*.

C- Pollen grain morphology (Table 5, Plate III)

The pollen grains considered small, as the polar axis never exceed 25 μm , irregular in shape, infolding, asymmetric and apolar. The shapes of the pollens are either subprolate in *Forsskaolea tenacissima* or prolate-spheroidal in *Parietaria alsinifolia*, *P.*

judaica, *Urtica pilulifera* and *U. urens* where P/E ratio was almost homogenous. The apertures are pantoaperturate, pantoporate with very small circular pores except in *Parietaria alsinifolia* and *P. judaica* they are panto-colporoidate with small pores. In *Urtica pilulifera* the pores are surrounded by smooth margo. The number of pores varies from 6-20 ectoexinous in all the studied species. The exine thin, with equal ectexine and endexine, except in *Urtica urens* the endexine is thicker than the ectexine. The ornamentation of the exine differs between the studied species, as it is tectate punctate in *Urtica pilulifera*, tectate granulate in *U. urens* or tectate verrucate in *Forsskaolea tenacissima*, *Parietaria alsinifolia* and *P. judaica*.

D- Seed morphology (Table 6, Plate IV)

The seed color is black and shiny in *Parietaria judaica*, off-white in *Urtica urens* and brown in *Forsskaolea tenacissima*, *P. alsinifolia* and *U. pilulifera*. The seed shape oblong ovate in *Forsskaolea tenacissima*, obovate in *Urtica urens* or ovate in *Parietaria alsinifolia*, *P. judaica* and *U. pilulifera*. The hilum basal in all studied species (depressed in *Urtica pilulifera* and *U. urens* and semi-depressed in *Forsskaolea tenacissima*, *Parietaria alsinifolia* and *P. judaica*). The surface sculpture rugose in *Forsskaolea tenacissima*, psilate in *Parietaria alsinifolia* and *P. judaica* or reticulate in *Urtica pilulifera* and *U. urens*. The epidermal cell shape polygonal in *Forsskaolea tenacissima*, irregular in *Urtica pilulifera* and *U. urens* or obscure in *Parietaria alsinifolia* and *P. judaica*. The anticlinal wall raised, straight shape and smooth surface in *Forsskaolea tenacissima*, *Urtica pilulifera* and *U. urens* or obscure in elevation, shape and surface in *Parietaria alsinifolia* and *P. judaica*. The periclinal wall surface granulate in *Forsskaolea tenacissima*, smooth in *Urtica pilulifera* and *U. urens* or obscure in *Parietaria alsinifolia* and *P. judaica*.

Systematic study on some Urticaceae Juss. species from Egypt

Table 3. Stem and lamina anatomical characters of the studied species of Urticaceae.

Taxa No.	Stem anatomical characters										Lamina anatomical characters			
	Out line in T.S.	Dermal System		Ground System					Vascular tissue		Epidermis shape	Mechanical tissue	Cystolith	Vascular bundles
		Cuticle	Epidermal Cells shape	Cortex				Stem internal appearance	Aspect	Interfascicular regions				
				Angular collenchyma	Chlorenchyma	Parenchyma	Extra-xylary Fibers							
1	Terete	Thick	Radially	+	-	+	+	Solid	Continuous siphonostele 19-20 bundles	Xylary fibers	Radially	-	+	One
2	Terete	Thin	Tangentially	-	-	+	-	Hollow	Continuous siphonostele 10-12 bundles	Xylary fibers	Tangentially	-	-	One
3	Terete	Thin	Tangentially	+	-	+	+	Solid	Continuous siphonostele 10-12 bundles	Xylary fibers	Radially	+	-	One
4	Ridged& furrowed	Thick	Tangentially	+	-	+	-	Hollow	Dissected siphonostele 10-12 bundles	Parenchyma	Radially	+	-	Three
5	Ridged& furrowed	Thick	Radially	+	+	+	-	Solid	Dissected siphonostele 10-12 bundles	Parenchyma	Tangentially	+	+	One

(+)= present, (-)= absent, 1: *Forsskaolea tenacissima*, 2: *Parietaria alsinifolia*, 3: *P. judaica*, 4: *Urtica pilulifera*, 5: *U. urens*

Table 4. Lamina abaxial epidermal characters of the studied species of Urticaceae.

Character Taxa	LM			SEM			
	Anticlinal wall shape	Sand Crystals	Trichomes	Surface sculpture	Anticlinal wall Elevation	Periclinal wall	
						Elevation	Surface
1	Undulate	Absent	E-glandular unicellular	Ruminant	Raised	Sunken	Granulate
2	Sinuate	Present	E-glandular unicellular	Rugose	Sunken	Raised	Striate
3	Sinuate	Absent	E-glandular unicellular	Ruminant	Raised	Sunken	Smooth
4	Undulate	Present	E-glandular unicellular& stinging hairs	Colliculate	Sunken	Raised	Smooth
5	Undulate	Present	E-glandular unicellular& stinging hairs	Colliculate	Sunken	Raised	Striate

1: *Forsskaolea tenacissima*, 2: *Parietaria alsinifolia*, 3: *P. judaica*, 4: *Urtica pilulifera*, 5: *U. urens*

Table 5. Quantitative and qualitative pollen morphological characters of the studied species of Urticaceae.

Character Taxa	Dimension	Pollen size	Shape class as shown by SEM	Apertures			Exine	
				Type	Number	Margo	Ectexine / Endexine	Sculpture
1	L= 12.97 µm W= 10.35 µm	Small	Subprolate	Pantoporate	10- 20	Absent	Equal	Verrucate
2	L= 15.14 µm W= 13.66 µm	Small	Prolate - spheroidal	Panto-colporoidate	6- 10	Absent	Equal	Verrucate
3	L= 10.66 µm W= 10.13 µm	Small	Prolate - spheroidal	Panto-colporoidate	6- 10	Absent	Equal	Verrucate
4	L= 22.86 µm W= 21.49 µm	Medium	Prolate - spheroidal	Pantoporate	8- 16	Present	Equal	Punctuate
5	L= 8.84 µm W= 8.57 µm	Small	Prolate - spheroidal	pantoporate	10- 20	Absent	Thicker endexine	Granulate

(L)= length instead of polar axis; (W)= width instead of equatorial diameter. 1: *Forsskaolea tenacissima*, 2: *Parietaria alsinifolia*, 3: *P. judaica*, 4: *Urtica pilulifera*, 5: *U. urens*

Table 6. Quantitative and qualitative seed morphological characters of the studied species of Urticaceae.

Character Taxa	Color	Shape	Dimension (L x W) mm	Hilum level	Surface sculpture	Epidermal cell shape	Anticlinal wall			Periclinal wall	
							Elevation	Shape	Surface	Elevation	Surface
1	Brown	Oblong ovate	8-10 x 7-9	Semi- depressed	Rugose	Polygonal	Raised	Straight	Smooth	Depressed	Granulate
2	Brown	Ovate	9-11 x 5-7	Semi- depressed	Psilate	Obscure	Obscure	Obscure	Obscure	Obscure	Obscure
3	Black	Ovate	9-11 x 6-8	Semi- depressed	Psilate	Obscure	Obscure	Obscure	Obscure	Obscure	Obscure
4	Brown	Obovate	10-12 x 8-10	Depressed	Reticulate	Irregular	Raised	Straight	Smooth	Depressed	Smooth
5	Off-white	Ovate	9-11 x 5-7	Depressed	Reticulate	Irregular	Raised	Straight	Smooth	Depressed	Smooth

(L)= length, (W)= width. 1: *Forsskaolea tenacissima*, 2: *Parietaria alsinifolia*, 3: *P. judaica*, 4: *Urtica pilulifera*, 5: *U. urens*

Systematic study on some Urticaceae Juss. species from Egypt

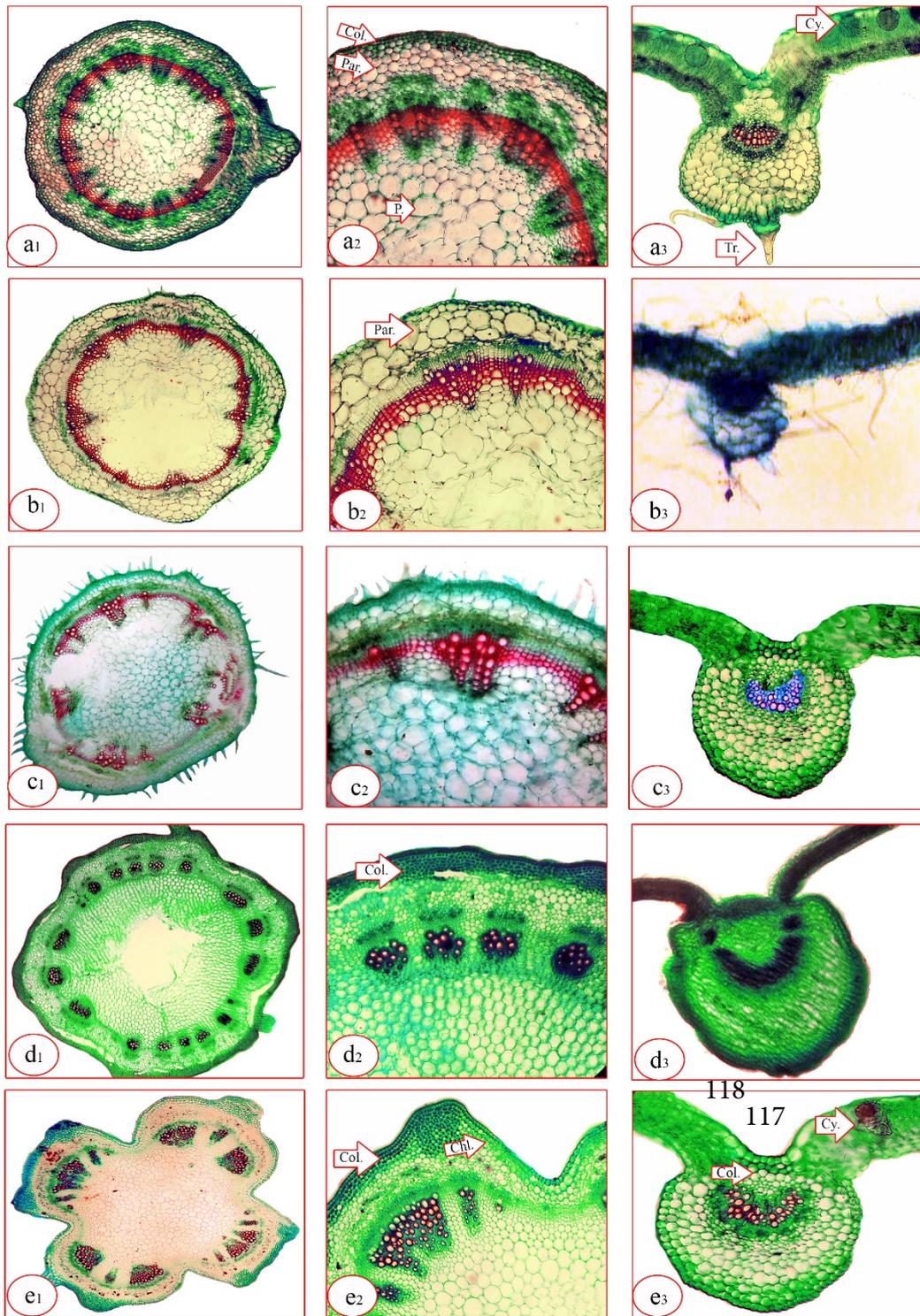


Plate I. Stem and lamina microphotographs showing different growth aspects of the studied species (LM, X= 20). a (1, 2 & 3): *Forsskaolea tenacissima*, b (1, 2 & 3): *Parietaria alsinifolia*, c (1, 2 & 3): *P. judaica*, d (1, 2 & 3): *Urtica pilulifera* & e (1, 2 & 3): *U. urens*. Chl.; chlorenchyma, Col.; collenchyma, Cy.; cystolith, P.; pith, Par.; parenchyma & Tr.; trichome.

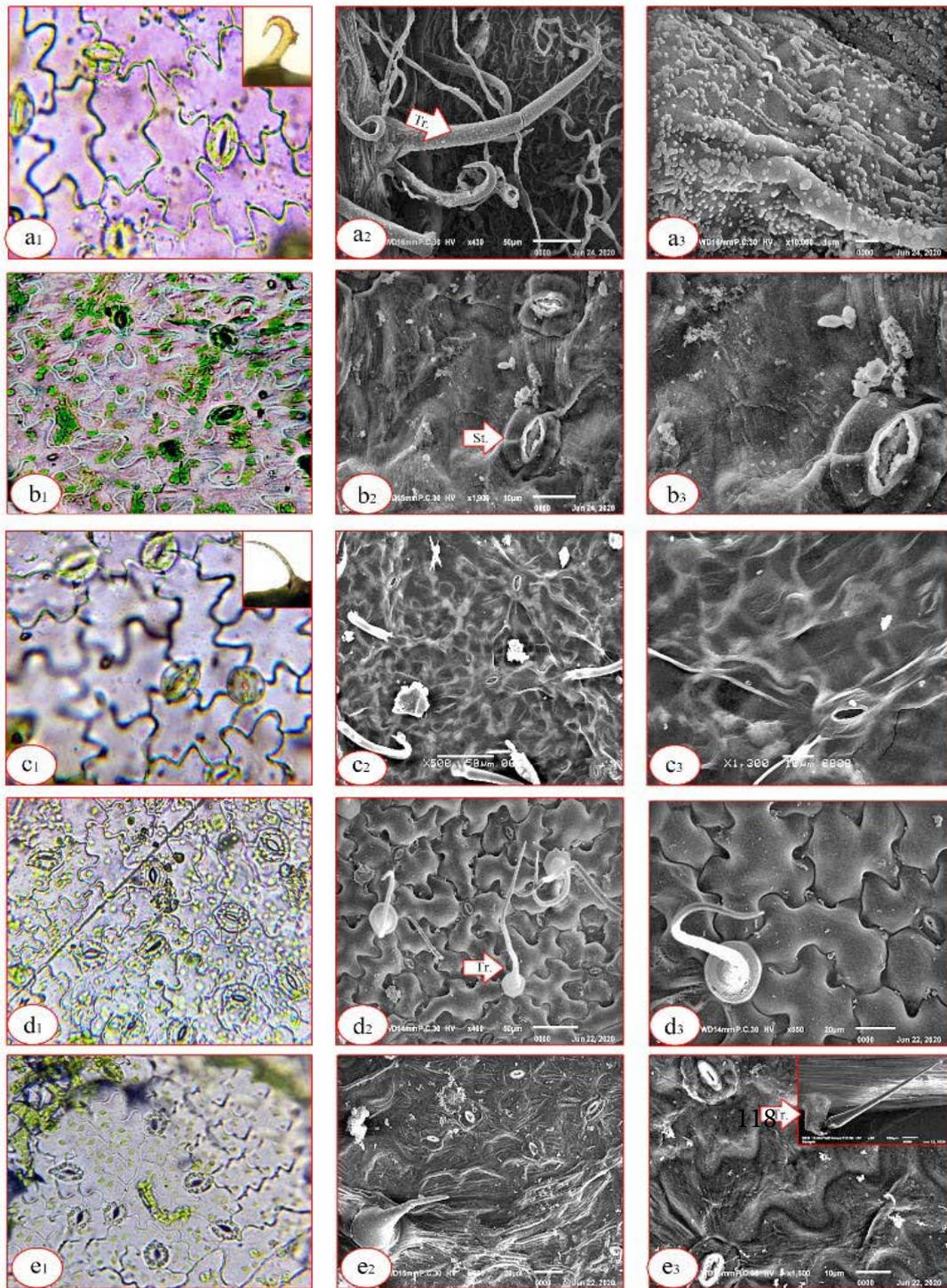


Plate II. Major aspects of lamina epidermal characters of the studied species (LM, X = 20 & SEM). a (1, 2 & 3): *Forsskaolea tenacissima*, b (1, 2 & 3): *Parietaria alsinifolia*, c (1, 2 & 3): *P. judaica*, d (1, 2 & 3): *Urtica pilulifera* & e (1, 2 & 3): *U. urens*. St.; stomata & Tr.; trichome.

Systematic study on some Urticaceae Juss. species from Egypt

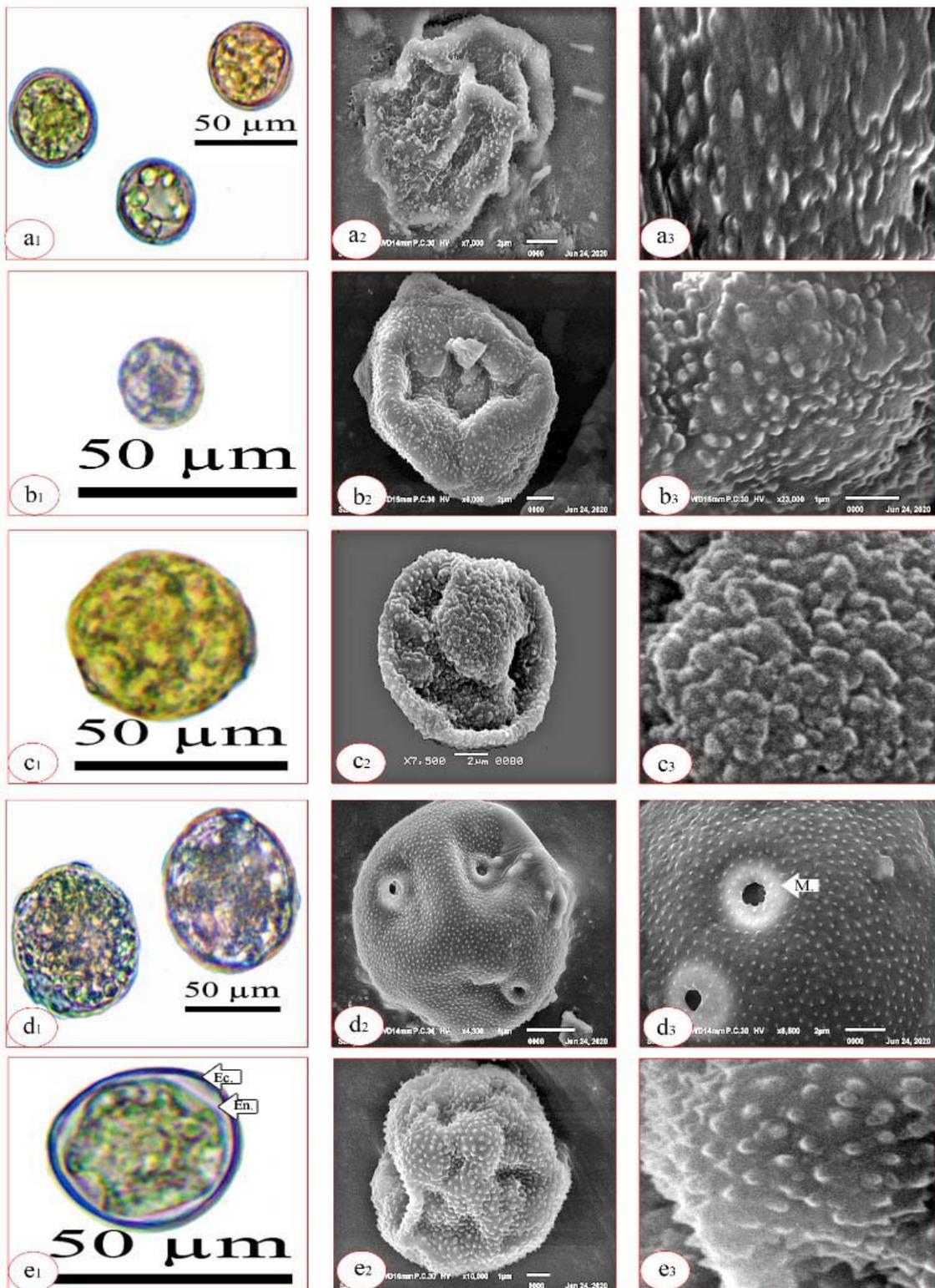


Plate III. Microphotographs of pollen morphology of the studied species (LM & SEM). a (1, 2 & 3): *Forsskaolea tenacissima*, b (1, 2 & 3): *Parietaria alsinifolia*, c (1, 2 & 3): *P. judaica*, d (1, 2 & 3): *Urtica pilulifera* & e (1, 2 & 3): *U. urens*. Ec; ectexine, En; endexine, M; margo

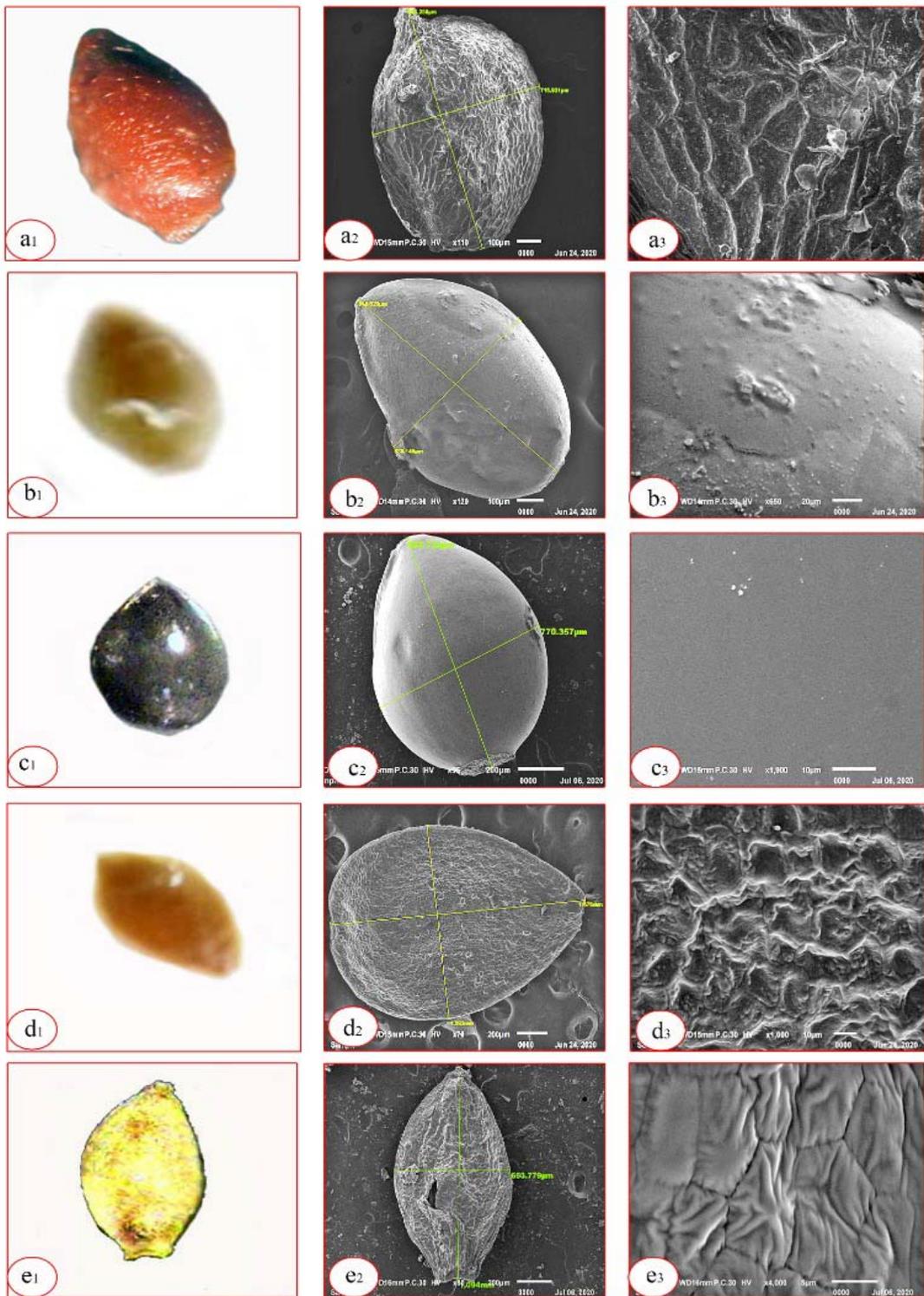


Plate IV. Microphotographs of seed morphology of the studied species (LM & SEM). a (1, 2 & 3): *Forsskaolea tenacissima*, b (1, 2 & 3): *Parietaria alsinifolia*, c (1, 2 & 3): *P. judaica*, d (1, 2 & 3): *Urtica pilulifera* & e (1, 2 & 3): *U. urens*.

Systematic study on some Urticaceae Juss. species from Egypt

An artificial key based on the most obvious characters is designed to facilitate the differentiation between the studied species as follow:

- 1a- Stem vascular tissue in form of dissected siphonostele..... 2
- 1b- Vascular tissue in form of continuous siphonostele.....3
- 2a- Three vascular bundle present in lamina midrib..... *Urtica pilulifera*
- 2b- One vascular bundle present in lamina midrib..... *Urtica urens*
- 3a- Seed coat ornamentation rugose..... *Forsskaolea tenacissima*
- 3b- Seed coat psilate.....4
- 4a- Seed testa black..... *Parietaria judaica*
- 4b- Seed testa brown..... *Parietaria alsinifolia*

E- Numerical analysis

The data were analyzed using the data matrix organized for five OTU's x 37 binary and multistate traits (Table 7). The obtained dendrogram is distinguished into

two clusters and three groups (Fig. 1). The plot of the five OTU's on the first three principal component axes is shown in figures 2 - 4.

Table 7. Characters, character states and their codes used in morphometric analysis of the studied species and the highest factor loading on the first three principal component.

No.	Character	Character state	1	2	3	4	5	PC 1	PC 2	PC 3	
								Loading factor			
1	Outline	Ridged & furrowed (0), Terete (1)	1	1	1	0	0	-0.161	-0.030777	0.066857	
2	Cuticle	Thick (0), Thin (1)	0	1	1	0	0	-0.089016	-0.19913	0.080171	
3	Epidermis	Tangential (0), Radial (1)	1	0	0	0	1	0.023482	0.21296	0.16746	
4	Cortex	Extra xylary fibers (0), Angular collenchyma (1)	1	0	1	1	1	0.052267	0.093342	0.061757	
5	Internal appearance	Hollow (0), Solid (1)	1	0	1	0	1	-0.013267	0.10718	0.30939	
6	Vascular tissue	Dissected siphonostele (0), Continuous siphonostele (1)	1	1	1	0	0	-0.161	-0.030777	0.066857	
7	Stem and lamina anatomy	Xylem in interfascular regions	1	1	1	0	0	-0.161	-0.030777	0.066857	
8		Lamina epidermis	Tangential (0), Radial (1)	0	1	0	0	1	0.043198	-0.04873	0.11902
9		Mechanical tissue	Absent (0), Present (1)	0	0	1	1	1	0.12425	-0.075008	0.075071
10	Lamina epidermis	Cystolith	1	0	0	0	1	0.023482	0.21296	0.16746	
11		No. of Vasular bundles	Three (0), One (1)	1	1	1	0	1	-0.065534	0.013835	0.24763
12		Anticlinal wall shape	Sinuate (0), Undulate (1)	1	0	0	1	1	0.089016	0.19913	-0.080171
13	Lamina epidermis	Sand crystals	0	1	0	1	1	0.10873	-0.062565	-0.12861	
14		Trichomes	E-glandular (0), E-glandular & stinging (1)	0	0	0	1	1	0.161	0.030777	-0.066857
15		Surface sculpture	Ruminate (0), Rugose (1), Colliculate (2)	0	1	0	2	2	0.26973	-0.031789	-0.19547
16		Anticlinal wall elevation	Sunken (0), Raised (1)	1	0	1	0	0	-0.10873	0.062565	0.12861

Mohamed A. Salim

17	Periclinal wall sculpture	Granulate (0), Striate (1), Smooth (2)	0	1	2	2	1	0.10077	-0.28797	-0.092387
18	Periclinal wall elevation	Sunken (0), Raised (1)	0	1	0	1	1	0.10873	-0.062565	-0.12861
19	Dimension (length x width; μm)	12.97 x 10.35 (0), 15.14 x 13.66 (1), 10.66 x 10.13 (2), 22.86 x 21.49 (3), 8.84 x 8.57 (4)	0	1	2	3	4	0.4527	-0.16797	0.2023
20	Size	Medium (0), Small (1)	1	1	1	0	1	-0.065534	0.013835	0.24763
21	Shape class	Prolate- spheroidal (0), Subprolate (1)	1	0	0	0	0	-0.071983	0.16835	-0.013314
22	Aperture type	Pantoporate (0), Panto-colporoidate (1)	0	1	1	0	0	-0.089016	-0.19913	0.080171
23	Aperture number	6-10 (0), 8-16 (1), 10-20 (2)	2	0	0	1	2	0.1125	0.41209	0.087287
24	Aperture margo	Absent (0), Present (1)	0	0	0	1	0	0.065534	-0.013835	-0.24763
25	Ectexine/ endexine thickness	Equal (0), Thicker (1)	0	0	0	0	1	0.095465	0.044612	0.18077
26	Exine sculpture	Verrucate (0), Punctate (1), Granulate (2)	0	0	0	1	2	0.25646	0.075388	0.11391
27	Dimension (length x width; mm)	8-10 x 7-9 (0), 9-11 x 5-7 (1), 9-11 x 6-8 (2), 10-12 x 8-10 (3), 9-11 x 5-7 (4)	0	1	2	3	4	0.4527	-0.16797	0.2023
28	Seed color	Brown (0), Black (1), Off-white (2)	0	0	1	0	2	0.15418	-0.016561	0.50347
29	Seed shape	Oblong ovate (0), Ovate (1), Obovate (2)	0	1	1	2	1	0.13752	-0.18218	-0.23432
30	Hilum position	Semi-depressed (0), Depressed (1)	1	1	1	0	0	-0.161	-0.030777	0.066857
31	Surface sculpture	Rugose (0), Psilate (1), Reticulate (2)	0	1	1	2	2	0.23298	-0.13757	-0.053543
32	Epidermal cell shape	Polygonal (0), Irregular (1), Obscure (2)	0	2	2	1	1	-0.017032	-0.36748	0.093485
33	Anticlinal wall elevation	Obscure (0), Raised (1)	1	0	0	1	1	0.089016	0.19913	-0.080171
34	Anticlinal wall shape	Obscure (0), Straight (1)	1	0	0	1	1	0.089016	0.19913	-0.080171
35	Anticlinal wall surface sculpture	Obscure (0), Smooth (1)	1	0	0	1	1	0.089016	0.19913	-0.080171
36	Periclinal wall elevation	Obscure (0), Depressed (1)	1	0	0	1	1	0.089016	0.19913	-0.080171
37	Periclinal wall surface sculpture	Granulate (0), Smooth (1), Obscure (2)	1	2	2	0	0	-0.25001	-0.2299	0.14703
Eigenvalue of principle component								11.0268	4.95773	2.11626
Percentage variance of principle component								56.839	25.555	10.909
Percentage total variation for the first three principal component amounts								93.303		

Systematic study on some Urticaceae Juss. species from Egypt

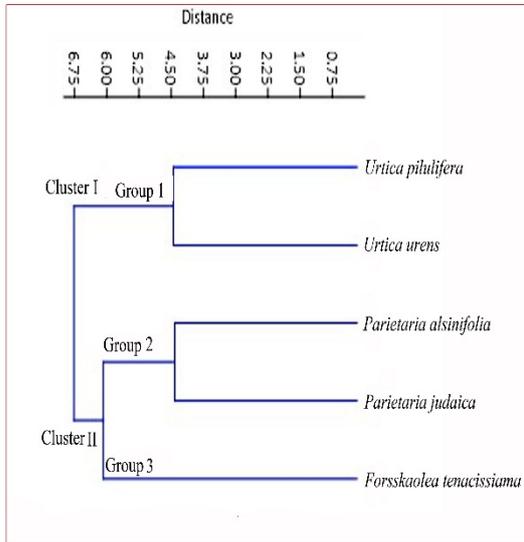


Fig. 1. Dendrogram based on 37 morphological characters of the studied species of Urticaceae.

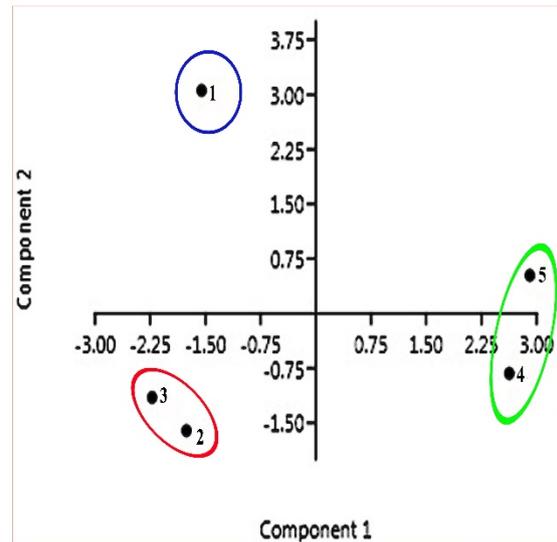


Fig. 2. Scatter plot of the 5 OTU's plotted against the first principal component by the second principal component.

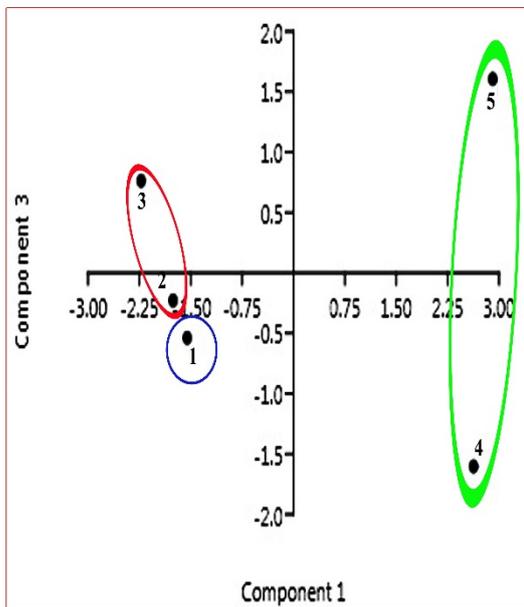


Fig. 3. Scatter plot of the 5 OTU's plotted against the first principal component by the third principal component.

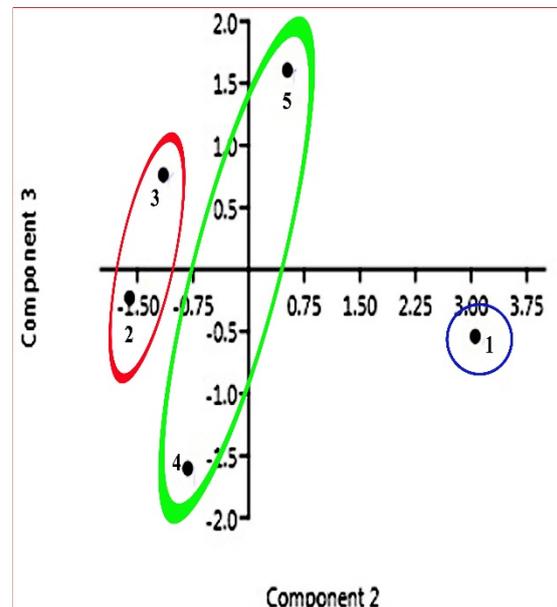


Fig. 4. Scatter plot of the 5 OTU's plotted against the second principal component by the third principal component.

Discussion

Family Urticaceae, which have been faced with many taxonomic opinions as shown in table 1, has been subjected in this study to reveal the dissimilarities between the different taxonomic categories indicated by Gaudichaud (1980), Weddell (1869), Friis (1993), Kravtsova (2009) and APG III (2009). In Egypt the family represented by three genera and six species according to Boulos (1999). These genera have been surveyed and examined externally and internally to clarify their affinities. The variable data obtained (37 character as shown in table 7) have been analyzed using PAST program version 3.16. The studied species has been divided into two clusters with three groups. The first cluster (group 1) gather *Urtica pilulifera* and *U. urens* at taxonomic distance 4.50. Cluster II subdivided into two groups (2 and 3). The former group includes *Parietaria alsinifolia* and *P. judaica* at taxonomic distance 4.50, while the second group has *Forsskaolea tenacissima* at taxonomic distance 6.00. These divisions separate the genera, each in a separate group with clear separations between the *Urtica* species than the two other genera. The studied *Urtica* species have specific characters which distinguished it from the two studied genera as the stem outline and the dissected siphonostelic structure without xylem fibers in between the vascular bundles at xylem region. *Urtica* species is the only one with stinging glandular trichomes and the seeds have depressed hilum. These notifications have been pointed before by Metcalfe & Chalk (1950), Jafari & Dehghan (2012) and Hamdy et al. (2016). The second cluster which separates the other two genera, each in a separate group. The first subgroup (group 2) has the two *Parietaria* species which distinguished from the *Forsskaolea tenacissima* (group 3) by having sinuate anticlinal epidermal cell walls, different type of pollen grain aperture, panto-colporoidate apertures and psilate seed coat surface. This separation of the two genera in accordance with seed

morphological characters obtained by Abid et al. (2015) who found that *Parietaria* and *Forsskaolea* species in the same phenetic clade and far from the *Urtica* species. As well as, this separation of the studied species coordinates with the obtained artificial key.

The plot of the five OTU's, on the first three principal components axes, shown in figures 2, 3 & 4 explain 93.303% of the total observed variation. On the first axis (56.839% total variance) a segregation is demonstrated between the three groups and the main characters explaining this separation (characters with high factor loading ≥ 0.16) are stem outline, the stem vascular tissue, component of xylem in interfascicular region, trichomes on lamina surface, lamina epidermal surface sculpture, pollen dimension, exine sculpture, seed hilum position, seed surface sculpture and periclinal wall sculpture. The second principal component axis (25.555% total variance) reveals splitting of the three groups based on stem epidermal cell shape in cross section, cuticle thickness, presence of cystolith in lamina wing, lamina anticlinal walls shape, lamina epidermal periclinal walls sculpture, pollen shape class, aperture type, number of pores per grain, seed shape, seed epidermal cell shape, the anticlinal and the periclinal walls. Along the third axis (10.909% total variance) the separation is based on stem epidermal cell shape in cross section, stem internal appearance, presence of cystolith, number of vascular bundles in midrib region in vertical section of lamina, lamina epidermal surface sculpture, pollen size, aperture margo, ectexine/ endexine thickness, seed shape and color. The most specific characters of different organs viz. stem, lamina, pollen and seed characters which facilitate the separation or grouping of the studied species in two clusters and

Systematic study on some Urticaceae Juss. species from Egypt

three groups are in accord with Metcalfe & Chalk (1950), Jafari & Dehghan (2012), Abid et al. (2015) and Hamdy et al. (2016). The obtained dendrogram (UPGMA) and principal component analysis (PCA) are similar in distinguishing the studied species into three groups, and this is in accord with Ghafoor (1981) and Abid et al. (2015) who splitted the taxa of Urticaceae based on seed morphological characters.

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Conclusion

The obtained data separates the three genera, each separately, according to their studied characters. This conclusion coordinate with the previous historic works on the family shown in table 1, as each of the studied genus has been classified in separate tribes. More phylogenetic works must be done for investigation the relationship between the Urticaceae taxa.

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